1	Microbial Systematics
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3	The Subdivision of the Genus <i>Eremothecium</i> Borzi emend. Kurtzman
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31	Keywords: Crebrothecium ashbyi; Nematospora coryli; Eremothecium cymbalariae;
32	Ashbya gossypii; Holleya sinecauda.
33	
34	Abstract
35 26	In the family Saccharomycetaceae, the genus <i>Eremothecium</i> Borzi emend. Kurtzman
36 37	included the five species that were on the whole characterized by the complex distribution of isoprepoid guipone homologues ( $0.5, 0.6, 0.7$ and $0.9$ ) as well as of peedle shaped
57	of isoprenoid quinone homologues (Q-5, Q-6, Q-7 and Q-9) as well as of needle-shaped

- 38 ascospore ornamentation. However, the calculated pair-wise sequence similarities among
- 39 the five species were very low (94.7 96.5%) in the 26S rRNA gene D1/D2 domain
- 40 sequences and so on. The experimental data obtained indicated that the emended genus
- 41 was not taxonomic homogeneous- but heterogeneous-natured, indicating that the five

species should be accommodated to their own separate five genera as *Eremothecium cymbalariae*, *Nematospora coryli*, *Ashbya gossypii*, *Crebrothecium ashbyi* and *Holleya sinecauda*.

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The genus *Holleya* was introduced on the basis of the exceptional distribution of ubiquinone-9 (Q-9) as respiratory quinone with the needle-shaped ascospore-forming species, *Holleya sinecaauda* (= *Nematospora sinecauda* Holley) (Yamada 1986).

50

51 In the phylogenetic analysis based on the partial base sequencing (Yamada and 52 Nagahama 1991), Holleya sinecauda (Q-9) represented one base substitution in positions 53 1685 - 1835, 151 bases of 26S rRNA, but did considerable base substitutions, i.e., seven 54 bases, when compared with Nematospora coryli (Q-6 or Q-5, Yamada et al. 1981) in 55 positions 1451 - 1618, 168 bases of 18S rRNA, demonstrating that the genus Holleya was 56 not similar phylogenetically but separable from the genus Nematospora. Incidentally, the 57 base substitutions were only four between Nematospora corvli and Saccharomyces 58 cerevisiae but eight between Holleya sinecauda and Saccharomyces cerevisiae. 59

Kurtzman (1995) compared the genera *Ashbya*, *Eremothecium*, *Holleya* and *Nematospora* by use of the 580 base-sequences near the 5'end of 26S rDNA. The
experimental results showed that the five species of the four genera were closely related
and the taxa was little divergence. As a conclusion, all the species concerned were placed
in the single genus *Eremothecium* Borzi emend. Kurtzman.

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66 This paper deals with the presently available sequence data and gives the different 67 conclusion that the five species are not closely related but phylogenetically independent 68 from one another to constitute their own taxonomic homogeneous-natured genera, as 69 previously reported (Yamada 2023).

70

71 The phylogenetic tree based on the 26S rRNA gene D1/D2 domain sequences (LSU 72 D1/D2) was constructed by the neighbour-joining method for the five species. As shown in Fig. 1, the phylogenetic branches of the five species were somewhat shorter than that 73 74 (94.0%) between Vanderwaltozyma polyspora and Saccharomyces cerevisiae in the family 75 Sacharomycetaceae (Table 1). However, the branch lengths were almost the same as that 76 (95.7%) between *Dipodascopsis uninucleata* and *Lipomvces starkevi* in the family 77 Lipomycetaceae used as reference standards. Among the five species, the calculated pair-78 wise sequence similarities were 94.7 - 96.5% and below 98% (Table 1). 79 80 For example, the calculated sequence similarity between Holleva sinecauda and

81 *Eremothecium cymbalariae* was 95.9%, which was almost the same as that (95.7%)

82 between Dipodascopsis uninucleata and Lipomyces starkeyi (Yamada et al. 2022). In

83 addition, the similarity (96.5%) between Holleva sinecauda and Nematospora corvli was

84 lower than that (97.5%) between Kawasakia arxii and Lipomyces starkeyi. The results

85 obtained above represented that the five species within the genus *Eremothecium* emend.

should be unequivocally divided into five genera respectively (Table 1).

- 86
- 87

88 In the phylogenetic tree based on the 18S rRNA gene sequences (SSU) derived from 89 the neighbour-joining method for the five species (Fig. 2), the phylogenetic branches of 90 the five species were on the whole short, when compared with those of the phylogenetic 91 tree based on LSU D1/D2. The pair-wise sequence similarities were high (98.4 - 99.4%) 92 (Table 2). On the other hand, the sequence similarity between Vanderwaltozyma polyspora 93 and Saccharomyces cerevisiae utilized as reference standards was also high as well 94 (98.8%). The phylogenetic data obtained above suggested that the base substitutions in 95 SSU were abnormally slow when compared with those of LSU, and such a curious nature 96 appeared to be distributed widely in the members of the family Saccharomycetaceae, since 97 the reference standards also showed very high sequence similarity (98.8%) (Table 2). The 98 two experimental data indicated that the five species should be classified in the separate 99 five genera.

100

101 The phylogenetic tree based on the concatenated sequences from the 18S rRNA genes 102 and 26S rRNA gene D1/D2 domains was constructed by the neighbour-joining method for 103 the five species. As shown in Fig. 3, the phylogenetic branches of the five species were longer when compared with those of SSU. The calculated pair-wise sequence similarities 104 105 were 97.6 - 98.4% (Table 3). On the other hand, the sequence similarity between Vanderwaltozyma polyspora and Saccharomyces cerevisiae utilized as reference standards gave a 106 107 similar sequence similarity (97.6%). The two experimental data suggested that the five 108 species should be classified in the separate five genera.

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110 Kurtzman and de Hoog (2011) represented the phylogenetic tree of the genus 111 Eremothecium emend. Kurtzman based on the concatenated gene sequences from LSU 112 rRNA, SSU rRNA, 5.85/alignable ITS rRNA, mitochondrial SSU rDNA, cytochrome 113 oxidase II and translation elongation factor- $1-\alpha$ . Since there was no information about the total number of used bases, it was temporally designated as 5500. The phylogenetic 114 branch-lengths were measured by use of a ruler, and the pair-wise sequence similarities 115 116 were calculated among the five species. The calculated results gave almost the same 117 sequence similarity, i.e., 93.8 - 96.7% and of course below 98% (Table 4). 118 As described above, the five species assigned to the genus Eremothecium emend. were 119

120 adequate to be subdivided into five genera respectively. Chemotaxonomically and

morphologically, the five species were very complicated in contrast to other members (Q-121

122 6) of the family Saccharomycetaceae (Yamada et al. 1977, 1981, 1987; Kurtzman 1995).

123	To distinguish the taxonomic small and unique group, the subfamily Eremothecioideae
124	was appropriately given.
125	
126	The subfamily Eremothecioideae subfam. nov., the family Saccharomycetaceae
127	
128	Genus I Eremothecium Borzi MB1883
129	Eremothecium cymbalariae Borzi (1888) MB235811
130	Q-7(Q-6)-equipped.
131	
132	Genus II Nematospora Peglion MB3441
133	Nematospora coryli Peglion (1897) MB222583
134	Synonym: Eremothecium coryli (Peglion) Kurtzman (1995)
135	Q-6-equipped (Q-5 only in the type strain).
136	
137	Genus III Ashbya Guillielmond MB389
138	Ashbya gossypii (Ashby et Nowell) Guillielmond (1928) MB266255
139	Synonym: Eremothecium gossypii (Ashby et Nowell) Kurtzman (1995)
140	Q-6-equipped.
141	
142	Genus IV Crebrothecium Routien MB1283
143	Crebrothecium ashbyi Routien (1949) MB266255
144	Synonym: Eremothecium ashbyi (Guillielmond) Kurtzman (1995)
145	Q-6(Q-7)-equipped.
146	
147	Genus V Holleya Yamada MB25105
148	Holleya sinecauda (Holley) Yamada (1986) MB131133
149	Synonym: Eremothecium sinecaudum (Holley) Kurtzman (1995)
150	Q-9(Q-8)-equipped.
151	
152	The genus Crebrothecium Routien was synonymous with the genus Eremothecium
153	Borzi (Kurtzman and de Hoog 2011). However, the present experimental data has shown
154	that it is phylogenetically deniable, since the calculated sequence similarity was low but
155	not high (96.5%, Table 1; 95.9%., Table 4; 96.6%, Yamada 2023).
156	
157	The five species classified in the subfamily Eremothecioideae were quite unique,
158	differing from other members (Q-6) of the family Saccharomycetaceae. The complex
159	distribution of isoprenoid quinone homologues as well as needle-shaped ascospore
160	ornamentation in the genus Eremothecium emend. was reasonable, since the emended
161	genus was a taxonomic heterogeneous-natured taxon.
162	
163	Conclusions

4

164	Kurtzman (1995, 2003) and Kurtzman and Robnett (1998) should notice the branch
165	lengths (= the so-called evolutionary distances) in the phylogenetic trees in the generic
166	designation, namely, the longer the branches were, the more taxonomic heterogeneous-
167	natured genera would be born.
168	
169	According to the present authors' experiences, the calculated sequence similarities
170	have to be 98% or more (beyond the so-called 98% barrier) in the species concerned to
171	constitute a taxonomic homogeneous-natured genus (Yamada et al. 2022; Huong et al.
172	2022a, b; Yamada 2023).
173	
174	The experimental data obtained above indicated that the present authors' generic
175	concept, i.e., the existence of taxonomic homogeneous-natured genera was phylo-
176	genetically and taxonomically reasonable.
177	
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186	The authors declare that there are no conflicts of interest.
187	
188	Author contributions
189	T. M., H.T.L. V., P. Y., S. T. and Y. Y. designed the study. T. M. performed the main
190	experiments. H.T.L. V. and P. Y. instructed the experiments. Y. Y. prepared the manuscript.
191	The detailed discussions were made among the five.
192	
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Table 1. The pair-wise sequence similarity in the 26S rRNA gene D1/D2 domain sequences in the five species.

Species	1	2	3	4	5	6	7
1. Crebrothecium ashbyi	100						
2. Nematospora coryli	94.7	100					
3. Eremothecium cymbalaraie	96.5	96.3	100				
4. Ashbya gossipii	94.7	95.8	96.1	100			
5. Holleya sinecauda	95.2	96.5	95.9	94.9	100		
6. Saccharomyces cerevisiae	86.9	85.4	86.6	86.9	85.4	100	
7. Vanderaltozyma polyspora	87.5	85.9	87.1	86.8	86.4	94.0	100
Species	8	9	10	11			
8. Lipomyces starkaeyi	100						
9. Waltomyces lipofer	95.2	100					
10. Dipodascopsis uninucleata	95.7	94.0	100				
11. Kawasakia arxii	97.5	94.1	95.4	100			

The percent similarity was calculated by use of 573 - 630 bases. All the strains used were the type strains or the authentic strains (See Fig. 1).

Table 2. The pair-wise sequence similarity in the 18S rRNA gene sequences in the five species.

	•		-	-		-	
Species	1	2	3	4	5	6	7
1. Crebrothecium ashbyi	100						
Species Species	9 <mark>8</mark> .6	100 2		3	4		5
1.3 Eremothecium cymbalaraie	986	99.0	100				
2 <sup>4</sup> Ashbya gossipii 2. Nematospora coryli	<u>88:8</u>	99.4 <mark>100</mark>	99.1	100			
3.5 <sub>E</sub> Holleya sinecauda 3. Eremothecium cymbalaraie	<u> 98:4</u>	99. <mark>96.3</mark>	98.7	$100^{99.1}$	100		
4. Ashbya gossipii	<u> 86:9</u>	97. <mark>9</mark> 5.8	97.0	96.1 <sup>97.5</sup>	97 <mark>100</mark>	100	
5 <sup>7</sup> Vanderaltozyma polyspora	<u>86.7</u>	96.96.5	96.8	95 <sup>97.1</sup>	96 <mark>7</mark> 9	98.8	100
Species	8	9 7	10	8 <sup>11</sup>	9		
6. Lipomyces starkaeyi	188						
79 Waltomyces lipofer 7. Waltomyces lipofer	<u>85:4</u>	100 <sub>100</sub>					
10 Dipodascopsis uninucleata 8. Dipodascopsis uninucleata	<u>95:</u> 9	97.5 <mark>.94.0</mark>	100	100			
11, Kawasakia arxii 9. Kawasakia arxii	<u> 86.9</u>	96.84.1	97.5	95.4 <sup>100</sup>	100		

The percent similarity was calculated by use of 1746 - 1790 bases. All the strains used were the type strains or the authentic strains (See Fig. 2).

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Table 3. The pair-wise sequence similarity in the concatenated sequences from the 18S rRNA genes and 26S rRNA gene D1/D2 domains in the five species.

Species	1	2	3	4	5	6	7
1. Crebrothecium ashbyi	100						
2. Nematospora coryli	97.6	100					
3. Eremothecium cymbalaraie	98.1	98.3	100				
4. Ashbya gossipii	97.8	98.4	98.4	100			
5. Holleya sinecauda	97.6	98.5	98.0	98.1	100		
6. Saccharomyces cerevisiae	94.5	94.3	94.5	94.8	94.2	100	
7. Vanderaltozyma polyspora	94.6	94.3	94.6	94.7	94.4	97.6	100
Species	8	9	10	11			
8. Lipomyces starkaeyi	100						
9. Waltomyces lipofer	95.4	100					
10. Dipodascopsis uninucleata	95.6	96.6	100				
11. Kawasakia arxii	96.3	96.1	97.0	100			

The percent similarity was calculated by use of 2278 - 2280 bases. All the strains used were the type strains or the authentic strains (See Fig. 3).

Table 4 The pair-wise sequence similarity in the five species

Species 1 2 3 4 5								
Species	1	2	3	4	5			
1. Ashbya gossypii	100							
2. Crebrothecium ashbyi	96.3	100						
3. Eremothaecium cymbalariae	96.1	95.9	100					
4. Nematospora cryli	95.1	95.4	96.7	100				
5. Holleya sinecauda	93.8	94.1	95.4	96.5	100			

Data was cited from the phylogenetic tree based on the concatenated sequences from LSU, SSU,

ITS, mitochondrial SSU, cytochrome oxidase II and elongation factor- $1\alpha$  (Kurtzman and de Hoog 2011). The percent similarities were calculated by use of the total 5500 bases.

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266 1 2 3 4 Species 5 267 100 268 1. Ashbya gossypii 2. Crebrothecium ashbyi 96.3 100 96.1 8 95.9 3. Eremothaecium cymbalariae 100 95.1 4. Nematospora cryli 95.4 96.7 100 5. Holleya sinecauda 93.8 94.1 95.4 96.5 100





0.02 K<sub>nuc</sub>.

Fig. 1. The phylogenetic tree based on the 26S rRNA gene D1/D2 domain sequences with 559 bases for the five species derived from the neighbour-joining method. The numerals at the nodes of the respective branches indicate bootstrap values (%) deduced from 1000 replications. 





0.01 K<sub>nuc</sub>.

Fig. 2. The phylogenetic tree based on the 18S rRNA gene sequences with 1661 bases for the five species derived from the neighbour-joining method. The numerals at the nodes of the respective branches indicate bootstrap values (%) deduced from 1000 replications.



0.01 K<sub>nuc</sub>.

Fig. 3. The phylogenetic tree based on the concatenated sequences from the 18S rRNA genes and 26S rRNA gene D1/D2 domains with 2220 bases for the five species derived from the neighbour-joining method. The numerals at the nodes of the respective branches indicate bootstrap values (%) deduced from 1000 replications.