

1 Microbial Systematics (Short Communication)

2  
3 **The Generic Circumscription of *Mrakia* and Related Taxa (Psychrophilic Yeasts)**

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27 **Keywords:** *Mrakia*; *Mrakia frigida*; *Mrakiella*; *Mrakiella cryoconiti*; *Krasilnikovozyma*  
28 *curviuscula* comb. nov..

29  
30 **Abstract**

31 In the family Mrakiaceae, the type genus *Mrakia* sensu stricto included five teleomorphic  
32 species with the type species, *Mrakia frigida*. In contrast, the anamorphic genus *Mrakiella*  
33 sensu stricto did 11 species with the type species, *Mrakiella cryoconiti*. Between the two  
34 genera, the completely separated clusters were shown in the phylogenetic tree (LSU  
35 D1/D2) derived from the maximum parsimony method. The pair-wise sequence similarity  
36 between *Mrakia frigida* and *Mrakiella cryoconiti* and *Mrakiella aquatica* were 98.2% and  
37 96.8% respectively. The calculated pair-wise sequence similarities were 100-99.3% among  
38 the five *Mrakia* species and were 97.1% between *Mrakiella cryoconiti* and *Mrakiella*  
39 *aquatica*. The teleomorphic genus *Krasilnikovozyma* emend. contained three species with  
40 the type species, *Krasilnikovozyma curviuscula*. Thus, the two teleomorphic genera were  
41 taxonomic homogeneous-natured, and the three were characteristic of Q-8.

<p>The family Mrakiaceae Liu et al.</p> <p>Genus</p> <p><i>Mrakia</i> Yamada et Komagata sensu stricto</p> <p><i>Mrakiella</i> Margesin et Fell sensu stricto</p> <p><i>Krasilnikovozyma</i> Liu et al. emend.</p>	<p>Species</p> <p><i>Mrakia frigida</i><sup>T</sup> (Fell et al.) Yamada et Komagata (1987)</p> <p><i>Mrakia gelida</i> (Fell et al.) Yamada et Komagata (1987)</p> <p><i>Mrakia psychrophila</i> Xin et Zhou (2007)</p> <p><i>Mrakia robertii</i> Thomas-Hall et Turchetti (2010)</p> <p><i>Mrakia blollopsis</i> Thomas-Hall et Turchetti (2010)</p> <p><i>Mrakiella crioconiti</i><sup>T</sup> Margesin et Fell (2008)</p> <p><i>Mrakiella aquatica</i> (Jones et Slooff) Margesin et Fell (2008)</p> <p><i>Mrakiella nicombsii</i> Thomas-Hall (2010)</p> <p><i>Mrakiella arctica</i> (Tsuji) comb. nov.</p> <p><i>Mrakiella hoshinonis</i> (Tsuji et al.) comb. nov.</p> <p><i>Mrakiella fibulata</i> (Yurkov et Turchetti) comb. nov.</p> <p><i>Mrakiella panshiensis</i> (Jia et Hui) comb. nov.</p> <p><i>Mrakiella stelviica</i> (Turchetti et Buzzini) comb. nov.</p> <p><i>Mrakiella montana</i> (Turchetti et Buzzini) comb. nov.</p> <p><i>Mrakiella terrae</i> (Park et al.) comb. nov.</p> <p><i>Mrakiella soli</i> (Park et al.) comb. nov.</p> <p><i>Krasilnikovozyma curviuscula</i><sup>T</sup> (Bav'eva et al.) comb. nov.</p> <p><i>Krasilnikovozyma huempii</i> f.a. (Ramirez et Gonzalez) Liu et al. (2015)</p> <p><i>Krasilnikovozyma tahquamenonensis</i> f.a. (Wang et al.) Liu et al. (2015)</p>
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<p>The family Mrakiaceae Liu et al.</p> <p>Genus</p> <p><i>Mrakia</i> Yamada et Komagata emend. Liu et al.</p> <p><i>Krasilnikovozyma</i> Liu et al.</p>	<p>Species</p> <p><i>Mrakia frigida</i><sup>T</sup> (Fell et al.) Yamada et Komagata (1987)</p> <p><i>Mrakia gelida</i> (Fell et al.) Yamada et Komagata (1987)</p> <p><i>Mrakia psychrophila</i> Xin et Zhou (2007)</p> <p><i>Mrakia robertii</i> Thomas-Hall et Turchetti (2010)</p> <p><i>Mrakia blollopsis</i> Thomas-Hall et Turchetti (2010)</p> <p><i>Mrakia crioconiti</i> (Margesin et Fell) Liu et al. (2015)</p> <p><i>Mrakia aquatica</i> (Jones et Slooff) Liu et al. (2015)</p> <p><i>Mrakia nicombsii</i> (Thomas-Hall) Liu et al. (2015)</p> <p><i>Mrakia arctica</i> Tsuji (2017)</p> <p><i>Mrakia hoshinonis</i> Tsuji et al. (2019)</p> <p><i>Mrakia fibulata</i> Yurkov et Turchetti (2019)</p> <p><i>Mrakia panshiensis</i> Jia et Hui (2020)</p> <p><i>Mrakia stelviica</i> Turchetti et Buzzini (2020)</p> <p><i>Mrakia montana</i> Turchetti et Buzzini (2020)</p> <p><i>Mrakia terrae</i> Park et al. (2021)</p> <p><i>Mrakia soli</i> Park et al. (2021)</p> <p><i>Krasilnikovozyma huempii</i><sup>T</sup> (Ramirez et Gonzalez) Liu et al. (2015)</p> <p><i>Krasilnikovozyma tahquamenonensis</i> (Wang et al.) Liu et al. (2015)</p>
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45 The genus *Mrakia* Yamada et Komagata was separated from the genus *Leucospori-*  
46 *dium* Fell et al. and introduced with *Mrakia frigida* as the type species based on the  
47 characteristic isoprenoid quinone-8 (Q-8) (Yamada and Komagata 1987). Up to now, five  
48 species have been reported: *Mrakia frigida*, *Mrakia gelida*, *Mrakia psychrophila*, *Mrakia*  
49 *robertii* and *Mrakia bolollopsis*. In contrast, the anamorphic genus *Mrakiella* Margesin et  
50 Fell was proposed with the type species, *Mrakiella cryoconiti* (Margesin and Fell 2008),  
51 and the 11 species have been reported.

52 Later, the genus *Mrakiella* was transferred taxonomically to the teleomorphic genus  
53 *Mrakia* with emendation (Liu et al. 2015). The genus *Mrakia* Yamada et Komagata emend.  
54 Liu et al. formed the monophyletic group along with *Krasilnikovozyma*, *Phaffia*,  
55 *Udeniomyces*, *Itersoniella* and *Tausonia* (Liu et al. 2015).

56 This paper is concerned with the revival of the genus *Mrakiella* on the basis of the  
57 phylogenetic separation within the genus *Mrakia* emend. i.e., the teleomorphic and the  
58 anamorphic groups, the former of which was especially taxonomic homogeneous-natured  
59 again.

60

61 The family Mrakiaceae Liu, Bai, Groenew et Boekhout, the order Cystofilobasidiales  
62 Fell, Roeyman et Boekhout:

63

64 Genus I. *Mrakia* Yamada et Komagata sensu stricto (MB25264)

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66 One to three-celled metabasidium with basidiospores is shown (Fell 2011), extremely  
67 short phylogenetic branches are produced within the genus in a phylogenetic tree (LSU  
68 D1/D2) and ubiquinone-8

69 The type species is *Mrakia frigida*.

70

71 1. *Mrakia frigida* (Fell, Stanzell, Hunter et Phaff) Yamada et Komagata (1987)  
72 (MB135389)

73 Basionym: *Leucosporidium frigidum* Fell, Stanzell, Hunter et Phaff (1969)

74

75 2. *Mrakia gelida* (Fell, Stanzell, Hunter et Phaff) Yamada et Komagata (1987)  
76 (MB135390)

77 Basionym: *Leucosporidium gelidum* Fell, Stanzell, Hunter et Phaff (1969)

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79 3. *Mrakia psychrophila* Xin et Zhou (2007) (MB508500)

80

81 4. *Mrakia robertii* Thomas-Hall et Turchetti (2010) (MB514690)

82

83 5. *Mrakia blollopsis* Thomas-Hall et Turchetti (2010) (MB514691)

84

85 In the phylogenetic tree based on the 26S rRNA gene D1/D2 domain sequences, the  
86 clusters of the teleomorphic and the anamorphic species were completely separated from  
87 each other (Thomas-Hall et al. 2010). In addition, the phylogenetic branches of the five  
88 *Mrakia* species were abnormally short, when compared with those of the anamorphic  
89 representative species, *Mrakiella cryoconiti* and *Mrakiella aquatica*, suggesting that the  
90 appearance of *Mrakia* species on the earth was relatively new from the view-point of  
91 evolutionary aspect.

92 The pair-wise sequence similarities between the type species, *Mrakia frigida* and the  
93 remaining four *Mrakia* species were extremely high (100 - 99.5%) (Table 1). In contrast,  
94 the sequence similarities between *Mrakia frigida* and *Mrakiella cryoconiti* and *Mrakiella*  
95 *aquatica* were low (98.2 and 96.8%). Among the five *Mrakia* species, the calculated pair-  
96 wise sequence similarities were 100 - 99.3% (data not shown).

97 To introduce the taxonomic homogeneous-natured genus, the calculated pair-wise  
98 sequence similarities were 98% or more between *Kockiozyma suominensis* and *Myxozyma*  
99 *geophila* (= *Kockiozyma geophila* f.a.; Lipomycetaceae) (Yamada et al. 2022) and between  
100 *Octosporomyces octosporus* (= *Schizosaccharomyces octosporus*) and *Octosporomyces*  
101 *osmophilus* (= *Schizosaccharomyces osmophilus*; Schizosaccharomycetaceae) (Vu et al.  
102 2022a) in the 26S rRNA gene D1/D2 domain sequences. In the 18S rRNA gene sequences,  
103 98% or more sequence similarities were also calculated to accomodate seven *Myxozyma*  
104 species to the teleomorphic genus *Kockiozyma* (Lipomycetaceae) (Vu et al. 2022b).

105 From the data obtained above, the teleomorphic genus *Mrakia* sensu stricto should be  
106 accepted, since the sequence similarities were extremely high (99.5% or more) in the  
107 family Mrakiaceae.

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109 Genus II. *Mrakiella* Margesin et Fell sensu stricto (MB536881)

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111 No metabasidium is shown (Fell and Margesin 2011), long phylogenetic branches are  
112 produced within the genus in a phylogenetic tree (LSU D1/D2) and ubiquinone-8

113 The type species is *Mrakiella crioconiti*

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115 1. *Mrakiella crioconiti* Margesin et Fell (2008) (MB537002)

116

117 2. *Mrakiella aquatica* (Jones et Slooff) Margesin et Fell (2008) (MB514705)

118 Basionym: *Candida aquatica* Jones et Slooff (1966)

119

120 3. *Mrakiella nicombsii* Thomas-Hall (2010) (MB514692)

121

122 4. *Mrakiella arctica* (Tsuji) comb. nov.

123 Basionym: *Mrakia arctica* Tsuji, Mycoscience, 59: 57 (2017) (MB821502)

124 The type strain is JCM32070<sup>T</sup>

125

- 126 5. *Mrakiella hoshinonis* (Tsuji, Tanabe, Vincent et Uchida) comb. nov.  
127 Basionym: *Mrakia hoshinonis* Tsuji, Tanabe, Vincent et Uchida, Int. J. Syst. Evol.  
128 Microbiol., DOI 10.1099/ijsem.0.003216, 4 (2019) (MB825484)  
129 The type strain is JCM 32575<sup>T</sup>.  
130
- 131 6. *Mrakiella fibulata* (Yurkov et Turchetti) comb. nov.  
132 Basionym: *Mrakia fibulata* Yurkov et Turchetti, Antonie van Leeuwenhoek,  
133 <https://doi.org/10.1007/s10482-019-01359-4>, 8 (2019) (MB 830398)  
134 The type strain is DSM 103931<sup>T</sup>.  
135
- 136 7. *Mrakiella panshiensis* (Jia et Hui) comb. nov.  
137 Basionym: *Mrakia panshiensis* Jia et Hui, Mycokeys, 74: 82 (2020) (MB834813)  
138 The type strain is NYNU 18562<sup>T</sup>.  
139
- 140 8. *Mrakiella stelviica* (Turchetti et Buzzini) comb. nov.  
141 Basionym: *Mrakia stelviica* Turchetti et Buzzini, Int. J. Syst. Evol. Microbiol. 70:  
142 4707 (2020) (MB835624)  
143 The type strain is DBVPG10734<sup>T</sup>  
144
- 145 9. *Mrakiella montana* (Turchetti et Buzzini) comb. nov.  
146 Basionym: *Mrakia montana* Turchetti et Buzzini, Int. J. Syst. Evol. Microbiol. 70:  
147 4709 (2020) (MB835626)  
148 The type strain is CBS 16462<sup>T</sup>.  
149
- 150 10. *Mrakiella terrae* (Park, Maeng et Sathiyaraj) comb. nov.  
151 Basionym: *Mrakia terrae* Park, Maeng et Sathiyaraj, Mycobiology, 49: 470 (2021)  
152 (MB836844)  
153 The type strain is YP416<sup>T</sup>.  
154
- 155 11. *Mrakiella soli* (Park, Maeng et Sathiyaraj) comb. nov.  
156 Basionym: *Mrakia soli* Park, Maeng et Sathiyaraj, Mycobiology, 49: 472 (2021)  
157 (MB836847)  
158 The type strain is YP421<sup>T</sup>.  
159
- 160 In contrast to the teleomorphic species of the genus *Mrakia*, the anamorphic  
161 *Mrakiella* species represented relatively long phylogenetic branches, indicating that the  
162 evolutionary stages may be distinct from one another. Within the genus *Mrakiella*, there is  
163 none of teleomorphic species, being different from the relationship between *Myxozyma*  
164 and *Kockiozyma* species (Lipomycetaceae) (Yamada et al. 2022).

165 The calculated pair-wise sequence similarities within the genus *Mrakiella* were quite  
166 diverse (97.1 - 98.8%) (Table 1), as observed in the genera *Myxozyma*, *Candida* and  
167 *Cryptococcus*.

168 According to Turchetii et al. (2020), *Mrakia stelviica* and *Mrakia montana* produced  
169 basidiospores from germinating teliospores and to Zhang et al. (2020), *Mrakia pan-*  
170 *schiensis* represented the teleomorphic stage, i.e., teliospores were produced and might  
171 germinate by a bud-like projection.

172

173 Genus III. *Krasilnikovozyma* Liu et al. emend. (MB812178)

174

175 Non-septate tubular metabasidium with sporidia is shown (Fell 2011), relatively short  
176 phylogenetic branches are produced within the genus in a phylogenetic tree (LSU D1/D2)  
177 and ubiquinone-8

178 The type species is *Krasilnikovozyma curviuscula*.

179

180 1. *Krasilnikovozyma curviuscula* (Bav'eva, Lisichkina, Reshetova et Danilevitch)  
181 comb. nov.

182 Basionym: *Mrakia curviuscula* Bav'eva, Lisichkina, Reshetova et Danilevitch,  
183 Mycobiology, 71: 450 (2002) (MB529873)

184 The type strain is CBS 9136<sup>T</sup>.

185

186 2. *Krasilnikovozyma huempii* f.a. (Ramirez et Gonzalez) Liu et al. (2015) (MB812179)

187

188 3. *Krasilnikovozyma tahquamenonensis* f.a. (Wang et al.) Liu et al. (2015)

189 (MB813656)

190

191 According to Fell (2011), *Mrakia curviuscula* (= *Krasilnikovozyma curviuscula*)  
192 produced a non-septate tubular metabasidium with one to two sporidia, which appeared to  
193 differ morphologically from those of *Mrakia frigida* and *Mrakia gelida*.

194 Liu et al. (2015) introduced the genus *Krasilnikovozyma* as an anamorphic taxon, since  
195 the type species was designated to be *Krasilnikovozyma huempii* (= *Cryptococcus*  
196 *huempii*). From the view-point of the traditional yeast systematics, it appeared to be  
197 problematic. Namely, the basic characteristics of living things on the earth are based on  
198 their reproduction, especially their sexual reproduction. Therefore, it is general that the  
199 teleomorphic genus has precedence over the anamorphic genus in the yeast systematics,  
200 and the name of the teleomorphic genus *Krasilnikovozyma* is able to be given to the  
201 anamorphic species (Lachance 2012).

202

203 In the phylogenetic tree based on the concatenated ITS and LSU D1/D2 sequences  
204 derived from the maximum likelihood method (Zhang et al. 2020), the cluster of the genus

205 *Mrakiella* was divided into two, i.e., one included *Mrakia stelviica* and *Mrakia montana*  
206 and the other did *Mrakia panshiensis*.

207 For the two teleomorphic species, *Mrakia stelviica* and *Mrakia montana* (Turchetti et al.  
208 2020), a new genus will be introduced, and for the one species, *Mrakia panshiensis*  
209 (Zhang et al. 2020), another new genus will be done.

210

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## 219 **Conflicts of interest**

220 The authors declare that there are no conflicts of interest.

221

## 222 **Author contributions**

223 Y.Y., H.T.L.V., P.Y. and S.T. designed the study. H.T.L.V. performed the main experiments.  
224 P.Y. instructed how to make the experiments. Y.Y. prepared the manuscript. The detailed  
225 discussions were made among Y.Y., H.T.L.V., P.Y., and S.T.

226

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241 philic, anamorphic, basidiomycetous yeast from alpine and arctic habitats. *Int J Syst*  
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260 genus for the Q<sub>8</sub>-equipped, self-sporulating organisms, which produce a unicellular  
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267 teleomorphic-stage of *M. arctica*. *Myckeys* **74**: 75-90.

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Table 1. The pair-wise sequence similarity of D1/D2 in *Mrakia* and *Mrakiella* species\*

Species	1	2	3	4	5	6	7
Sequence similarity (%)	100	100	99.8	99.5	99.5	98.2	96.8
Species	6	7	8	9	10		
Sequence similarity (%)	100	97.1	97.5	97.5	98.8		
Species	7	8	9	10			
Sequence similarity (%)	100	98.2	98.8	97.1			

\*The original data (the number of base substitution) was cited from Tsuji et al. (2019). In this case, the precise length of D1/D2 was not known. It was designated as 560 bases in sequence calculation.

1. *Mrakia frigida* CBS 5270<sup>T</sup>, 2. *Mrakia gelida* CBS 5272<sup>T</sup>, 3. *Mrakia robertii* 8912<sup>T</sup>, 4. *Mrakia blollopsis* CBS 8921<sup>T</sup>, 5. *Mrakia psychrophila* CBS 10829<sup>T</sup>, 6. *Mrakiella cryoconiti* CBS 10834<sup>T</sup>, 7. *Mrakiella aquatica* CBS 5443<sup>T</sup>, 8. *Mrakiella niccombsii* CBS 8917<sup>T</sup>, 9. *Mrakiella hoshinonis* JCM 32575<sup>T</sup>, 10. *Mrakiella arctica* JCM 32070<sup>T</sup>.

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