

1 Microbial Systematics (Short Communication)

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3 **The Generic Circumscription of *Mrakia* and Related Taxa (Psychrophilic Yeasts)**

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27 **Keywords:** *Mrakia*; *Mrakia frigida*; *Mrakiella*; *Mrakiella cryoconiti*; *Krasilnikovozyma*
28 *curviuscula* comb. nov..

29

30 **Abstract**

31 In the family Mrakiaceae, the type genus *Mrakia* sensu stricto included the five
32 teleomorphic species with the type species, *Mrakia frigida*. In contrast, the anamorphic
33 genus *Mrakiella* sensu stricto did the 11 species with the type species, *Mrakiella*
34 *cryoconiti*. Between the two genera, the completely separated clusters were shown in the
35 phylogenetic tree (LSU D1/D2) derived from the MP method. The pair-wise sequence
36 similarity between *Mrakia frigida* and *Mrakiella cryoconiti* and *Mrakiella aquatica* were
37 98.2% and 96.8%. The calculated pair-wise sequence similarities were 100-99.3% among
38 the five *Mrakia* species and were 97.1% between *Mrakiella cryoconiti* and *Mrakiella*
39 *aquatica*. The teleomorphic genus *Krasilnikovozyma* emend. contained the three species
40 with the type species, *Krasilnikovozyma curviuscula*. Thus, the two teleomorphic genera
41 were taxonomic homogeneous-natured, and the three were characteristic of Q-8.

43 **Supplementary Abstract**

The family Mrakiaceae Liu et al.	
Genus	Species
<i>Mrakia</i> Yamada et Komagata sensu stricto	<i>Mrakia frigida</i> ^T (Fell et al.) Yamada et Komagata (1987) <i>Mrakia gelida</i> (Fell et al.) Yamada et Komagata (1987) <i>Mrakia psychrophila</i> Xin et Zhou (2007) <i>Mrakia robertii</i> Thomas-Hall et Turchetti (2010) <i>Mrakia blolopsis</i> Thomas-Hall et Turchetti (2010) <i>Mrakiella crioconiti</i> ^T Margesin et Fell (2008) <i>Mrakiella aquatica</i> (Jones et Slooff) Margesin et Fell (2008) <i>Mrakiella nicombsii</i> Thomas-Hall (2010) <i>Mrakiella arctica</i> (Tsuiji) comb. nov. <i>Mrakiella hoshinonis</i> (Tsuiji et al.) comb. nov. <i>Mrakiella fibulata</i> (Yukov et Turchetti) comb. nov. <i>Mrakiella panshiensis</i> (Jia et Hui) comb. nov. <i>Mrakiella stelviica</i> (Turchetti et Buzzini) comb. nov. <i>Mrakiella montana</i> (Turchetti et Buzzini) comb. nov. <i>Mrakiella terrae</i> (Park et al.) comb. nov. <i>Mrakiella soli</i> (Park et al.) comb. nov.
<i>Krasilnikovozyma</i> Liu et al. emend.	<i>Krasilnikovozyma curviuscula</i> ^T (Bavjeva et al.) comb. nov. <i>Krasilnikovozyma huempii</i> f.a. (Ramirez et Gonzalez) Liu et al. (2015) <i>Krasilnikovozyma tahquamenonensis</i> f.a. (Wang et al.) Liu et al. (2015)

The family Mrakiaceae Liu et al.	
Genus	Species
<i>Mrakia</i> Yamada et Komagata emend. Liu et al.	<i>Mrakia frigida</i> ^T (Fell et al.) Yamada et Komagata (1987) <i>Mrakia gelida</i> (Fell et al.) Yamada et Komagata (1987) <i>Mrakia psychrophila</i> Xin et Zhou (2007) <i>Mrakia robertii</i> Thomas-Hall et Turchetti (2010) <i>Mrakia blolopsis</i> Thomas-Hall et Turchetti (2010) <i>Mrakia crioconitin</i> (MARGESIN et FELL) LIU et al. (2015) <i>Mrakia aquatica</i> (JONES et SLOOFF) LIU et al. (2015) <i>Mrakia nicombsii</i> (THOMAS-HALL) LIU et al. (2015) <i>Mrakia arctica</i> TSUJI (2017) <i>Mrakia hoshinonis</i> TSUJI et al. (2019) <i>Mrakia fibulata</i> YUKOV et TURCHETTI (2019) <i>Mrakia panshiensis</i> JIA et HUI (2020) <i>Mrakia stelviica</i> TURCHETTI et BUZZINI (2020) <i>Mrakia montana</i> TURCHETTI et BUZZINI (2020) <i>Mrakia terrae</i> PARK et al. (2021) <i>Mrakia soli</i> PARK et al. (2021)
<i>Krasilnikovozyma</i> Liu et al.	<i>Krasilnikovozyma huempii</i> ^T (RAMIREZ et GONZALEZ) LIU et al. (2015) <i>Krasilnikovozyma tahquamenonensis</i> (WANG et al.) LIU et al. (2015)

45 The genus *Mrakia* Yamada et Komagata was separated from the genus *Leucosporidium*
46 Fell et al. and introduced with *Mrakia frigida* as the type species based on the
47 characteristic isoprenoid quinone-8 (Q-8) (Yamada and Komagata 1987). Up to now, five
48 species have been reported: *Mrakia frigida*, *Mrakia gelida*, *Mrakia psychrophila*, *Mrakia*
49 *robertii* and *Mrakia bolollopsis*. In contrast, the anamorphic genus *Mrakiella* Margesin et
50 Fell was proposed with the type species, *Mrakiella cryoconiti* (Margesin and Fell 2008),
51 and the 11 species have been reported.

52 Later, the genus *Mrakiella* was transferred taxonomically to the teleomorphic genus
53 *Mrakia* with emendation (Liu et al. 2015). The genus *Mrakia* Yamada et Komgata emend.
54 Liu et al. formed the monophyletic group along with *Krasilnikovozyma*, *Phaffia*,
55 *Udeniomyces*, *Itersonilia* and *Tausonia* (Liu et al. 2915).

56 This paper is concerned with the revival of the genus *Mrakiella* on the basis of the
57 phylogenetic separation within the genus *Mrakia* emend. i.e., the teleomorphic and the
58 anamorphic groups, the former of which was especially taxonomic homogeneous-natured
59 again.

60
61 The family Mrakiaceae Liu, Bai, Groenew et Boekhout, the order Cystofilobasidiales
62 Fell, Roeijman et Boekhout:

63
64 Genus I. *Mrakia* Yamada et Komagata sensu stricto (MB25264)

65
66 One to three-celled metabasidium with basidiospores is shown (Fell 2011), extremely
67 short phylogenetic branches are produced within the genus in a phylogenetic tree (LSU
68 D1/D2) and ubiqinone-8

69 The type species is *Mrakia frigida*.

70
71 1. *Mrakia frigida* (Fell, Statzell, Hunter et Phaff) Yamada et Komagata (1987)
72 (MB135389)

73 Basionym: *Leucosporidium frigidum* Fell, Statzell, Hunter et Phaff (1969)

74
75 2. *Mrakia gelida* (Fell, Statzell, Hunter et Phaff) Yamada et Komagata (1987)
76 (MB135390)

77 Basionym: *Leucosporidium gelidum* Fell, Statzell, Hunter et Phaff (1969)

78
79 3. *Mrakia psychrophila* Xin et Zhou (2007) (MB508500)

80
81 4. *Mrakia robertii* Thomas-Hall et Turchetti (2010) (MB514690)

82
83 5. *Mrakia bolollopsis* Thomas-Hall et Turchetti (2010) (MB514691)

85 In the phylogenetic tree based on the 26S rRNA gene D1/D2 domain sequences, the
86 clusters of the teleomorphic and the anamorphic species were completely separated from
87 each other (Thomas-Hall et al. 2010). In addition, the phylogenetic branches of the five
88 *Mrakia* species were abnormally short, when compared with those of the anamorphic
89 representative species, *Mrakiella cryoconiti* and *Mrakiella aquatica*, suggesting that the
90 appearance of *Mrakia* species on the earth was relatively new from the view-point of
91 evolutionary aspect.

92 The pair-wise sequence similarities between the type species, *Mrakia frigida* and the
93 remaining four *Mrakia* species were extremely high (100 - 99.5%) (Table 1). In contrast,
94 the sequence similarities between *Mrakia frigida* and *Mrakiella cryoconiti* and *Mrakiella*
95 *aquatica* were low (98.2 and 96.8%). Among the five *Mrakia* species, the calculated pair-
96 wise sequence similarities were 100 - 99.3% (data not shown).

97 To introduce the taxonomic homogeneous-natured genus, the calculated pair-wise
98 sequence similarities were 98% or more between *Kockiozyma suominensis* and *Myxozyma*
99 *geophila* (= *Kockiozyma geophila* f.a.; Lipomycetaceae) (Yamada et al. 2022) and between
100 *Octosporomyces octosporus* (= *Schizosaccharomyces octosporus*) and *Octosporomyces*
101 *osmophilus* (= *Schizosaccharomyces osmophilus*; Schizosaccharomycetaceae) (Vu et al.
102 2022a) in the 26S rRNA gene D1/D2 domain sequences. In the 18S rRNA gene sequences,
103 98% or more sequence similarities were also calculated to accommodate seven *Myxozyma*
104 species to the teleomorphic genus *Kockiozyma* (Lipomycetaceae) (Vu et al. 2022b).

105 From the data obtained above, the teleomorphic genus *Mrakia* sensu stricto should be
106 accepted, since the sequence similarities were extremely high (99.5% or more) in the
107 family Mrakiaceae.

109 Genus II. *Mrakiella* Margesin et Fell sensu stricto (MB536881)

111 No metabasidium is shown (Fell and Margesin 2011), long phylogenetic branches are
112 produced within the genus in a phylogenetic tree (LSU D1/D2) and ubiquinone-8

113 The type species is *Mrakiella crioconiti*

115 1. *Mrakiella crioconiti* Margesin et Fell (2008) (MB537002)

117 2. *Mrakiella aquatica* (Jones et Slooff) Margesin et Fell (2008) (MB514705)

118 Basionym: *Candida aquatica* Jones et Slooff (1966)

120 3. *Mrakiella nicombsii* Thomas-Hall (2010) (MB514692)

122 4. *Mrakiella arctica* (Tsuiji, Tanabe, Vincent et Uchida) comb. nov.

123 Basionym: *Mrakia arctica* Tsuiji, Tanabe, Vincent et Uchida (2017) (MB821502)

124 The type strain is JCM32070^T

- 126 5. *Mrakiella hoshinonis* (Tsugi, Tanabe, Vincent et Uchida) comb. nov.
127 Basionym: *Mrakia hoshinonis* Tsugi, Tanabe, Vincent et Uchida (2019) (MB825484)
128 The type strain is JCM 32575^T.
129
130 6. *Mrakiella fibulata* (Yuekov et Turchetti) comb. nov.
131 Basionym: *Mrakia fibulata* Yuekov et Turchetti (2019) (MB 830398)
132 The type strain is DSM 103931^T.
133
134 7. *Mrakiella panshiensis* (Jia et Hui) comb. nov.
135 Basionym: *Mrakia panshiensis* Jia et Hui (2020) (MB834813)
136 The type strain is NYNU 18562^T.
137
138 8. *Mrakiella stelviica* (Turchetti et Buzzini) comb. nov.
139 Basionym: *Mrakia stelviica* Turchetti et Buzzini (2020) (MB835624)
140 The type strain is DBVPG10734^T
141
142 9. *Mrakiella montana* (Turchetti et Buzzini) comb. nov.
143 Basionym: *Mrakia montana* Turchetti et Buzzini (2020) (MB835626)
144 The type strain is CBS 16462^T.
145
146 10. *Mrakiella terrae* (Park, Maeng et Sathiyaraj) comb. nov.
147 Basionym: *Mrakia terrae* Park, Maeng et Sathiyaraj (2021) (MB836844)
148 The type strain is YP416^T.
149
150 11. *Mrakiella soli* (Park, Maeng et Sathiyaraj) comb. nov.
151 Basionym: *Mrakia soli* Park, Maeng et Sathiyaraj (2021) (MB836847)
152 The type strain is YP421^T.
153
154 In contrast to the teleomorphic species of the genus *Mrakia*, the anamorphic
155 *Mrakiella* species represented relatively long phylogenetic branches, indicating that the
156 evolutionary stages may be distinct from one another. Within the genus *Mrakiella*, there is
157 none of teleomorphic species, being different from the relationship between *Myxozyma*
158 and *Kockiozyma* species (Lipomycetaceae) (Yamada et al. 2022).
159 The calculated pair-wise sequence similarities within the genus *Mrakiella* were quite
160 diverse (97.1 - 98.8%) (Table 1), as observed in the genera *Myxozyma*, *Candida* and
161 *Cryptococcus*.
162 According to Turchetii et al. (2020), *Mrakia stelviica* and *Mrakia montana* produced
163 basidiospores from germinating teliospores and to Zhang et al. (2020), *Mrakia pan-*
164 *schiensis* represented the teleomorphic stage, i.e., teliospores were produced and might
165 germinate by a bud-like projection.
166

167 Genus III. *Krasilnikovozyma* Liu et al. emend. (MB812178)

168
169 Non-septate tubular metabasidium with sporidia is shown (Fell 2011), short
170 phylogenetic branches are produced within the genus in a phylogenetic tree (LSU D1/D2)
171 and ubiquinone-8

172 The type species is *Krasilnikovozyma curviuscula*.

173
174 1. *Krasilnikovozyma curviuscula* (Bavjeva et al.) comb. nov.

175 Basionym: *Mraka curviuscula* Bavjeva, Lisichkina, Reshetova et Danilevitch
176 (2002) (MB529873)

177 The type strain is CBS 9136^T.

178
179 2. *Krasilnikovozyma huempii* f.a. (Ramirez et Gonzalez) Liu et al. (2015) (MB812179)

180
181 3. *Krasilnikovozyma tahquamenonensis* f.a. (Wang et al.) Liu et al. (2015)

182 (MB813656)

183
184 According to Fell (2011), *Mrakia curviuscula* (= *Krasilnikovozyma curviuscula*)
185 produced a non-septate tubular metabasidium with one to two sporidia, which differed
186 morphologically from those of *Mrakia frigida* and *Mrakia gelida*.

187 Liu et al. (2015) introduced the genus *Krasilnikovozyma* as an anamorphic taxon, since
188 the type species was designated to be *Krasilnikovozyma huempii* (= *Cryptococcus*
189 *huempii*). From the view-point of the traditional yeast systematics, it appeared to be
190 problematic. Namely, the basic characteristics of living things on the earth are based on
191 their reproduction, especially their sexual reproduction. Therefore, it is general that the
192 teleomorphic genus has precedence over the anamorphic genus in the yeast systematics,
193 and the name of the teleomorphic genus *Krasilnikovozyma* is able to be given to the
194 anamorphic species (Lachance 2012).

195
196 In the phylogenetic tree based on the concatenated ITS and LSU D1/D2 sequences
197 derived from the maximum likelihood method (Zhang et al. 2020), the cluster of the genus
198 *Mrakiella* was divided into two, i.e., one included *Mrakia stelviica* and *Mrakia montana*
199 and the other did *Mrakia panshiensis*.

200 For the two teleomorphic species, *Mrakia stelviica* and *Mrakia montana* (Turchetii et al.
201 2020), a new genus will be introduced, and for the one species, *Mrakia panshiensis*
202 (Zhang et al. 2020), another new genus will be done.

203
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214
215 Author contributions
216 Y.Y., H.T.L.V., P.Y. and S.T. designed the study. H.T.L.V. performed the main experiments.
217 P.Y. instructed how to make the experiments. Y.Y. prepared the manuscript. The detailed
218 discussions were made among Y.Y., H.T.L.V., P.Y., and S.T.
219
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 253 genus for the Q₈-equipped, self-sporulating organisms, which produce a unicellular
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 259 new member of the Cystofilobasidiales from soil in China and description of the
 260 teleomorphic-stage of *M. arctica*. *Mycobanks* **74**: 75-90.
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 265
 266

Table 1. The pair-wise sequence similarity of D1/D2 in *Mrakia* and *Mrakiella* species*

Species	1	2	3	4	5	6	7
Sequence similarity (%)	100	100	99.8	99.5	99.5	98.2	96.8
Species	6	7	8	9	10		
Sequence similarity (%)	100	97.1	97.5	97.5	98.8		
Species	7	8	9	10			
Sequence similarity (%)	100	98.2	98.8	97.1			

*The original data (the number of base substitution) was cited from Tsuji et al. (2019). In this case, the precise length of D1/D2 was not known. It was designated as 560 bases in sequence calculation.
 1. *Mrakia frigida* CBS 5270^T, 2. *Mrakia gelida* CBS 5272^T, 3. *Mrakia robertii* 8912^T, 4. *Mrakia blollopsis* CBS 8921^T, 5. *Mrakia psychrophila* CBS 10829^T, 6. *Mrakiella cryoconiti* CBS 10834^T, 7. *Mrakiella aquatica* CBS 5443^T, 8. *Mrakiella niccombsii* CBS 8917^T, 9. *Mrakiella hoshinonis* JCM 32575^T, 10. *Mrakiella arctica* JCM 32070^T.

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