

1 Microbial Systematics (Short Communication)

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3 **The Generic Circumscription of *Mrakia* and Related Taxa (Psychrophilic Yeasts)**

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5 Yuzo Yamada,<sup>1,4,5,\*</sup> Huong Thi Lan Vu,<sup>2</sup> Pattaraporn Yukphan,<sup>1</sup> Somboon Tanasupawat,<sup>3</sup>

6  
7 <sup>1</sup>BIOTEC Culture Collection (BCC), National Center for Genetic Engineering and  
8 Biotechnology (BIOTEC), National Science and Technology Development Agency  
9 (NSTDA), 113 Thailand Science Park, Phaholyothin Road, Klong 1, Klong Luang,  
10 Pathumthani 12120, Thailand

11 <sup>2</sup>Department of Microbiology, Faculty of Biology and Biotechnology, University of  
12 Science, Vietnam National University-HCM City, 227 Nguyen Van Cu Street, Ward 4,  
13 District 5, Hochiminh City, Vietnam

14 <sup>3</sup>Department of Biochemistry and Microbiology, Faculty of Pharmaceutical Sciences,  
15 Chulalongkorn University, 254 Phayathai Road, Wangmai, Pathumwan, Bangkok 10330,  
16 Thailand

17 <sup>4</sup>JICA Senior Overseas Volunteer, Japan international Cooperation Agency, Shibuya-ku,  
18 Tokyo 151-8559. Japan

19 <sup>5</sup>Laboratory of Applied Microbiology (Professor Emeritus), Department of Agricultural  
20 Chemistry, Faculty of Agriculture, Shizuoka University, 836 Ohya, Suruga-ku, Shizuoka  
21 422-8529, Japan

22  
23 \*Corresponding author: Yuzo Yamada

24 E-mail: ymdy333@kdt.biglobe.ne.jp

25 ORCID, 0000-0002-2799-7045

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27 **Keywords:** *Mrakia*; *Mrakia frigida*; *Mrakiella*; *Mrakiella cryoconiti*; *Krasilnikovozyma*  
28 *curviuscula* comb. nov..

29  
30 **Abstract**

31 In the family Mrakiaceae, the type genus *Mrakia* sensu stricto included the five  
32 teleomorphic species with the type species, *Mrakia frigida*. In contrast, the anamorphic  
33 genus *Mrakiella* sensu stricto did the 11 species with the type species, *Mrakiella*  
34 *cryoconiti*. Between the two genera, the completely separated clusters were shown in the  
35 phylogenetic tree (LSU D1/D2) derived from the MP method. The pair-wise sequence  
36 similarity between *Mrakia frigida* and *Mrakiella cryoconite* and *Mrakiella aquatica* were  
37 98.2% and 96.8%. The calculated pair-wise sequence similarities were 100-99.3% among  
38 the five *Mrakia* species and were 97.1% between *Mrakiella cryoconiti* and *Mrakiella*  
39 *aquatica*. The teleomorphic genus *Krasilnikovozyma* emend. contained the three species  
40 with the type species, *Krasilnikovozyma curviuscula*. Thus, the two teleomorphic genera  
41 were taxonomic homogeneous-natured, and the three were characteristic of Q-8.



45 The genus *Mrakia* Yamada et Komagata was separated from the genus *Leucospori-*  
46 *dium* Fell et al. and introduced with *Mrakia frigida* as the type species based on the  
47 characteristic isoprenoid quinone-8 (Q-8) (Yamada and Komagata 1987). Up to now, five  
48 species have been reported: *Mrakia frigida*, *Mrakia gelida*, *Mrakia psychrophila*, *Mrakia*  
49 *robertii* and *Mrakia bololopsis*. In contrast, the anamorphic genus *Mrakiella* Margesin et  
50 Fell was proposed with the type species, *Mrakiella cryoconiti* (Margesin and Fell 2008),  
51 and the 10 species have been reported.

52 Later, the genus *Mrakiella* was transferred taxonomically to the teleomorphic genus  
53 *Mrakia* with emendation (Liu et al. 2015). The genus *Mrakia* Yamada et Komagata emend.  
54 Liu et al. formed the monophyletic group along with *Krasilnikovozyma*, *Phaffia*,  
55 *Udeniomyces*, *Itersoniella* and *Tausonia* (Liu et al. 2015).

56 This paper is concerned with the revival of the genus *Mrakiella* on the basis of the  
57 phylogenetic separation within the genus *Mrakia* emend. i.e., the teleomorphic and the  
58 anamorphic groups, the former of which was especially taxonomic homogeneous-natured  
59 again.

60

61 The family Mrakiaceae Liu, Bai, Groenew et Boekhout, the order Cystofilobasidiales  
62 Fell, Roeyman et Boekhout:

63

64 Genus I. *Mrakia* Yamada et Komagata sensu stricto (MB25264)

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66 One to three-celled metabasidium with basidiospores is shown (Fell 2011), extremely  
67 short phylogenetic branches are produced within the genus in a phylogenetic tree (LSU  
68 D1/D2) and ubiquinone-8

69 The type species is *Mrakia frigida*.

70

71 1. *Mrakia frigida* (Fell, Stetzell, Hunter et Phaff) Yamada et Komagata (1987)  
72 (MB135389)

73 Basionym: *Leucosporidium frigidum* Fell, Stetzell, Hunter et Phaff (1969)

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75 2. *Mrakia gelida* (Fell, Stetzell, Hunter et Phaff) Yamada et Komagata (1987)  
76 (MB135390)

77 Basionym: *Leucosporidium gelidum* Fell, Stetzell, Hunter et Phaff (1969)

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79 3. *Mrakia psychrophila* Xin et Zhou (2007) (MB508500)

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81 4. *Mrakia robertii* Thomas-Hall et Turchetti (2010) (MB514690)

82

83 5. *Mrakia blollopsis* Thomas-Hall et Turchetti (2010) (MB514691)

84

85 In the phylogenetic tree based on the 26S rRNA gene D1/D2 domain sequences, the  
86 clusters of the teleomorphic and the anamorphic species were completely separated from  
87 each other (Thomas-Hall et al. 2010). In addition, the phylogenetic branches of the five  
88 *Mrakia* species were abnormally short, when compared with those of the anamorphic  
89 representative species, *Mrakiella cryoconiti* and *Mrakiella aquatica*, suggesting that the  
90 appearance of *Mrakia* species on the earth was relatively new from the view-point of  
91 evolutionary aspect.

92 The pair-wise sequence similarities between the type species, *Mrakia frigida* and the  
93 remaining four *Mrakia* species were extremely high (100 - 99.5%) (Table 1). In contrast,  
94 the sequence similarities between *Mrakia frigida* and *Mrakiella cryoconiti* and *Mrakiella*  
95 *aquatica* were low (98.2 and 96.8%). Among the five *Mrakia* species, the calculated pair-  
96 wise sequence similarities were 100 - 99.3% (data not shown).

97 To introduce the taxonomic homogeneous-natured genus, the calculated pair-wise  
98 sequence similarities were 98% or more between *Kockiozyma suominensis* and *Myxozyma*  
99 *geophila* (= *Kockiozyma geophila* f.a.; Lipomycetaceae) (Yamada et al. 2022) and between  
100 *Octosporomyces octosporus* (= *Schizosaccharomyces octosporus*) and *Octosporomyces*  
101 *osmophilus* (= *Schizosaccharomyces osmophilus*; Schizosaccharomycetaceae) (Vu et al.  
102 2022a) in the 26S rRNA gene D1/D2 domain sequences. In the 18S rRNA gene sequences,  
103 98% or more sequence similarities were also calculated to accomodate seven *Myxozyma*  
104 species to the teleomorphic genus *Kockiozyma* (Lipomycetaceae) (Vu et al. 2022b).

105 From the data obtained above, the teleomorphic genus *Mrakia* sensu stricto should be  
106 accepted, since the sequence similarities were extremely high (99.5% or more) in the  
107 family Mrakiaceae.

108

109 Genus II. *Mrakiella* Margesin et Fell sensu stricto (MB536881)

110

111 No metabasidium is shown (Fell and Margesin 2011), long phylogenetic branches are  
112 produced within the genus in a phylogenetic tree (LSU D1/D2) and ubiquinone-8

113 The type species is *Mrakiella crioconiti*

114

115 1. *Mrakiella crioconiti* Margesin et Fell (2008) (MB537002)

116

117 2. *Mrakiella aquatica* (Jones et Slooff) Margesin et Fell (2008) (MB514705)

118 Basionym: *Candida aquatica* Jones et Slooff (1966)

119

120 3. *Mrakiella nicombsii* Thomas-Hall (2010) (MB514692)

121

122 4. *Mrakiella arctica* (Tsuji, Tanabe, Vincent et Uchida) comb. nov.

123 Basionym: *Mrakia arctica* Tsuji, Tanabe, Vincent et Uchida (2017) (MB821502)

124 The type strain is JCM32070<sup>T</sup>

125

- 126 5. *Mrakiella hoshinonis* (Tsuji, Tanabe, Vincent et Uchida) comb. nov.  
127 Basionym: *Mrakia hoshinonis* Tsuji, Tanabe, Vincent et Uchida (2019) (MB825484)  
128 The type strain is JCM 32575<sup>T</sup>.  
129
- 130 6. *Mrakiella fibulata* (Yuekov et Turchetti) comb. nov.  
131 Basionym: *Mrakia fibulata* Yuekov et Turchetti (2019) (MB 830398)  
132 The type strain is DSM 103931<sup>T</sup>.  
133
- 134 7. *Mrakiella panshiensis* (Jia et Hui) comb. nov.  
135 Basionym: *Mrakia panshiensis* Jia et Hui (2020) (MB834813)  
136 The type strain is NYNU 18562<sup>T</sup>.  
137
- 138 8. *Mrakiella stelviica* (Turchetti et Buzzini) comb. nov.  
139 Basionym: *Mrakia stelviica* Turchetti et Buzzini (2020) (MB835624)  
140 The type strain is DBVPG10734<sup>T</sup>  
141
- 142 9. *Mrakiella montana* (Turchetti et Buzzini) comb. nov.  
143 Basionym: *Mrakia montana* Turchetti et Buzzini (2020) (MB835626)  
144 The type strain is CBS 16462<sup>T</sup>.  
145
- 146 10. *Mrakiella terrae* (Park, Maeng et Sathiyaraj) comb. nov.  
147 Basionym: *Mrakia* Park, Maeng et Sathiyaraj (2021) (MB836844)  
148 The type strain is YP416<sup>T</sup>.  
149
- 150 11. *Mrakiella soli* (Park, Maeng et Sathiyaraj) comb. nov.  
151 Basionym: *Mrakia soli* Park, Maeng et Sathiyaraj (2021) (MB836847)  
152 The type strain is YP421<sup>T</sup>.  
153

154 In contrast to the teleomorphic species of the genus *Mrakia*, the anamorphic  
155 *Mrakiella* species represented relatively long phylogenetic branches, indicating that the  
156 evolutionary stages may be distinct from one another. Within the genus *Mrakiella*, there is  
157 none of teleomorphic species, being different from the relationship between *Myxozyma*  
158 and *Kockiozyma* species (Lipomycetaceae) (Yamada et al. 2022).

159 The calculated pair-wise sequence similarities within the genus *Mrakiella* were quite  
160 diverse (97.1 - 98.8%) (Table 1), as observed in the genera *Myxozyma*, *Candida* and  
161 *Cryptococcus*.

162 According to Turchetti et al. (2020), *Mrakia stelviica* and *Mrakia montana* produced  
163 basidiospores from germinating teliospores and to Zhang et al. (2020), *Mrakia pan-*  
164 *schiensis* represented the teleomorphic stage, i.e., teliospores were produced and might  
165 germinate by a bud-like projection.  
166

167 Genus III. *Krasilnikovozyma* Liu et al. emend. (MB812178)

168

169 Non-septate tubular metabasidium with sporidia is shown (Fell 2011), short  
170 phylogenetic branches are produced within the genus in a phylogenetic tree (LSU D1/D2)  
171 and ubiquinone-8

172 The type species is *Krasilnikovozyma curviuscula*.

173

174 1. *Krasilnikovozyma curviuscula* (Bavjeva et al.) comb. nov.

175 Basionym: *Mrakia curviuscula* Bavjeva, Lisichkina, Reshetova et Danilevitch

176 (2002) (MB529873)

177 The type strain is CBS 9136<sup>T</sup>.

178

179 2. *Krasilnikovozyma huempii* f.a. (Ramirez et Gonzalez) Liu et al. (2015) (MB812179)

180

181 3. *Krasilnikovozyma tahquamenonensis* f.a. (Wang et al.) Liu et al. (2015)

182 (MB813656)

183

184 According to Fell (2011), *Mrakia curviuscula* (= *Krasilnikovozyma curviuscula*)  
185 produced a non-septate tubular metabasidium with one to two sporidia, which differed  
186 morphologically from those of *Mrakia frigida* and *Mrakia gelida*.

187 Liu et al. (2015) introduced the genus *Krasilnikovozyma* as an anamorphic taxon, since  
188 the type species was designated to be *Krasilnikovozyma huempii* (= *Cryptococcus*  
189 *huempii*). From the view-point of the traditional yeast systematics, it appeared to be  
190 problematic. Namely, the basic characteristics of living things on the earth are based on  
191 their reproduction, especially their sexual reproduction. Therefore, it is general that the  
192 teleomorphic genus has precedence over the anamorphic genus in the yeast systematics,  
193 and the name of the teleomorphic genus *Krasilnikovozyma* is able to be given to the  
194 anamorphic species (Lachance 2012).

195

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## 204 **Conflicts of interest**

205 The authors declare that there are no conflicts of interest.

206

## 207 **Author contributions**

208 Y.Y., H.T.L.V., P.Y. and S.T. designed the study. H.T.L.V. performed the main experiments.  
209 P.Y. instructed how to make the experiments. Y.Y. prepared the manuscript. The detailed  
210 discussions were made among Y.Y., H.T.L.V., P.Y., and S.T.

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Table 1. The pair-wise sequence similarity of D1/D2 in *Mrakia* and *Mrakiella* species\*

Species	1	2	3	4	5	6	7
Sequence similarity (%)	100	100	99.8	99.5	99.5	98.2	96.8
Species	6	7	8	9	10		
Sequence similarity (%)	100	97.1	97.5	97.5	98.8		
Species	7	8	9	10			
Sequence similarity (%)	100	98.2	98.8	97.1			

\*The original data (the number of base substitution) was cited from Tsuji et al. (2019). In this case, the precise length of D1/D2 was not known. It was designated as 560 bases in sequence calculation.  
 1. *Mrakia frigida* CBS 5270<sup>T</sup>, 2. *Mrakia gelida* CBS 5272<sup>T</sup>, 3. *Mrakia robertii* 8912<sup>T</sup>, 4. *Mrakia blolopsis* CBS 8921<sup>T</sup>, 5. *Mrakia psychrophila* CBS 10829<sup>T</sup>, 6. *Mrakiella cryoconiti* CBS 10834<sup>T</sup>, 7. *Mrakiella aquatica* CBS 5443<sup>T</sup>, 8. *Mrakiella niccombsii* CBS 8917<sup>T</sup>, 9. *Mrakiella hoshinonis* JCM 32575<sup>T</sup>, 10. *Mrakiella arctica* JCM 32070<sup>T</sup>.

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